

SCAR markers linked with disease resistance traits in common bean (*Phaseolus vulgaris*)

Updated: May, 2009

SCAR Name	Marker of Origin	Pathogen	Size (bp) / orientation	Sequences of SCARS	Tagged Locus	LG	Reference
SAP6	AP6	Common Bacterial Blight (CBB)	820 cis	GTC ACG TCT CCT TAA TAG TA GTC ACG TCT CAA TAG GCA AA	Major QTL (GN#1 sel 27)	B10	Miklas et al., 2000b,c
BAC6	BC409	CBB	1250 cis	TAG GCG GCG GCG CAC GTT TTG TAG GCG GCG GAA GTG GCG GTG	Major QTL (GN#1 sel 27)	B10	Jung et al., 1999
SU91	U9	CBB	700 cis	CCA CAT CGG TTA ACA TGA GT CCA CAT CGG TGT CAA CGT GA	Major QTL (XAN 159)	B8	Pedraza et al., 1997
LG5 syn. BC420	BC420	CBB	900 cis	GCA GGG TTC GAA GAC ACA CTG G GCA GGG TTC GCC CAA TAA CG	Major QTL (XAN 159)	B6	Yu et al., 2000
R7313		CBB	700 cis	ATT GTT ATC GTC GAC ACG AAT ATT TCT GAT CAC ACG AG	Major QTL (OAC 88-1)	B8	Bai et al., 1997 Beattie et al., 1998
R4865		CBB	950 cis	TCC AAA GCC ATT CTA GTT CAG CTA CTT TCA AAC TGG G	Major QTL (OAC 88-1)		Bai et al., 1997 Beattie et al., 1998
SR13	R13	Halo bacterial blight (HBB)	1150 cis	GGA CGA CAA GGA ACA TAT TCA GGA CGA CAA GGC TGC AAG AAC CAT	<i>Pse-1</i>	B10	Miklas et al., 2009

ST8	T8	HBB	1350 cis	AAC GGC GAC ATC AGT GTA AAG G AAC GGC GAC AAC CGA CCA TGT TTT AC	<i>Pse-1</i>	B10	Miklas et al., 2009
SH11	H11	HBB	800 cis	CTT CCG CAG TCG AGA GAT CTT CCG CAG TAG CAC C	<i>Pse-1</i>	B10	Miklas et al., 2009
SB10	B10	HBB	525 cis	CTG CTG GGA CAA TCA CCA AGT C CTG CTG GGA CTC TCT TAC	<i>Pse-Unnamed</i>	B4	Fourie et al., 2004
SW13	W13	Bean Common Mosaic Virus (BCMV) & HBB	690 cis	CAC AGC GAC ATT AAT TTT CCT TTC CAC AGC GAC AGG AGG AGC TTA TTA	<i>I</i> <i>Pse-3</i>	B2	Haley et al., 1994 Melotto et al., 1996 Fourie et al., 2004
ROC11	C11	BCMV	420 trans	CCA ATT CTC TTT CAC TTG TAA CC GCA TGT TCC AGC AAA CC	<i>bc-3</i>	B6	Johnson et al., 1997
SBD5	BD5	BCMV	1250 cis	GTG CGG AGA GGC CAT CCA TTG GTG GTG CGG AGA GTT TCA GTG TTG ACA	<i>bc-1²</i>	B3	Miklas et al., 2000a

SR2	R2	Bean Golden Yellow Mosaic Virus (BGYMV)	530 / 570 codominant	CAC AGC TGC CCT AAC AAA AT CAC AGC TGC CAC AGG TGG GA	<i>bgm-1</i>	B3	Urrea et al., 1996 Beebe, PC, 1996 Blair et al., 2007
SW12	W12	BGYMV	700 cis	TGG GCA GAA GTT CTA GCA TGT GGC TGG GCA GAA GCA CAG TAT GAT TTG	Major QTL (DOR 364)	B4	Miklas et al., 2000c Singh et al. 2000
SAS8	AS08	Beet curly top virus (BCTV)	1550 cis	GGC TGC CAG TAT CTT GTC TAA CAC C GGC TGC CAG TGA CGC AAT TCT GCA G	<i>Bct</i>	B7	Larsen and Miklas, 2004
SK14	K14	Rust	620 cis	CCC GCT ACA CAC CAA TAC CTG CCC GCT ACA CTT GAT AAA ATG TTA G	<i>Ur-3</i>	B11*	Haley et al. 1994 Nemchin. & Stavelly, 1998 *Miklas et al., 2002
SA14	A14	Rust	1079 / 800 codominant	CTA TCT GCC ATT ATC AAC TCA AAC GTG CTG GGA AAC ATT ACC TAT T	<i>Ur-4</i>	B6*	Miklas et al. 1993 Miene et al. 2004 *Miklas et al., 2002
SI19	I19	Rust	460 cis	AAT GCG GGA GAT ATT AAA AGG AAA G AAT GCG GGA GTT CAA TAG AAA AAC C	<i>Ur-5</i>	B4*	Haley et al., 1993 Melotto et al., 1998 *Miklas et al., 2000c
SBC6	BC06	Rust	308 cis	GAA GGC GAG AAG AAA AAG AAA AAT GAA GGC GAG AGC ACC TAG CTG AAG	<i>Ur-6</i>	B11*	Park et al., 2003b, 2004b *Miklas et al. 2002

SAD12	AD12	Rust	537 cis	AAG AGG GCG TGA GAT CGT CG AAG AGG GCG TCT TGA AGG TT	<i>Ur-7</i>	B11	Park et al., 2003a, 2004a, 2008
SAE19	AE19	Rust	890 trans	CAG TCC CTG ACA ACA TAA CAC C CAG TCC CTA AAG TAG TTT GTC CCT A	<i>Ur-11</i>	B11*	Johnson et al., 1995 Queiroz et al., 2004c *Miklas et al., 2002
UR11-GT2	GT02	Rust	450 cis	CGC ACT TAG GAG CAC AAA TGG TGG GTC CCA TAT TTT G	<i>Ur-11</i>	B11*	Boone et al., 1999 *Miklas et al., 2002
KB126	E-AAC/M- ACC	Rust	405 / 430 codominant	GAA TTC AAC CTC GGC CAC TAC C TTA AAC CTT CCG GAG GAT TC	Ur-13	B8	Mienie et al., 2005
SF10	F10	Rust	1072 cis	GGA AGC TTG GTG AGC AAG GA GGA AGC TTG GCT ATGATG GT	Ouro Negro	B4*	Correa et al., 2000 *Miklas et al., 2002
SBA8	BA8	Rust	530 cis	CCA CAG CCG ACG GAG GAG GCC ATG TTT TTT GTC CCC	Ouro Negro	B4*	Correa et al., 2000 *Miklas et al., 2002
Phs	Phaseolin 'T' & 'S' alleles	White Mold & Common Bacterial Blight CBB	Multiple	AGC ATA TTC TAG AGG CCT CC GCT CAG TTC CTC AAT CTG TTC	Major QTL (G 122) & (BAT 93)	B7	Kami et al., 1995 Nodari et al., 1993 Miklas et al., 2001

SAU5	AU05	White mold	1350 cis	GAG CTA CCG TCA GTT TAC TAA GAG CTA CCG TGG CTT TTT TCT	QTL (minor) NY6020-4	B6	Miklas et al., 2003
SS18	S18	White mold	1650 cis	CTG GCG AAC TGT ACA TGC AAC ATA C CTG GCG AAC TGA TTC ATA CAT TTT G	QTL (major) NY6020-4	B8	Miklas et al., 2003
	SE _{ACT} /M _{CCA}	Anthracnose	codominant	AAT TCA CTT ATA AAA AAT AAA ATT AAC CAT AAC TGT TAT CAG ACC	<i>Co-1</i> ²	B1	Vallejo and Kelly, 2008
SCAreoli	H20	Anthracnose	1000	GGG AGA CAT CCA TCA GAC AAC TCC GTA TCC ATT TGAA GGA GCT	<i>Co-2</i>	B11	Geffroy et al., 1998 Adam-Blondon et al., 1994
SQ4	OQ4	Anthracnose & Rust	1440	CCT TAG GTA TGG TGG GAA ACG A TGA GGG CGA GGA TTT CAG CAA GTT	<i>Co-2</i> , <i>Ur-11</i>	B11	Awale et al., 2008 Young and Kelly, 1996
SW12	W12	Anthracnose	700 cis	TGG GCA GAA GTT CTA GCA TGT GGC TGG GCA GAA GCA CAG TAT GAT TTG	Co-3 /Co-9	B4	Miklas et al., 2000c Singh et al. 2000 Rodríguez-Suárez et al., 2008

SY20	Y20	Anthravnose	830 cis	AGC CGT GGA AGG TTG TCA T CCG TGG AAA CAA CAC ACA AT	<i>Co-4</i>	B8*	Queiroz et al., 2004b *Kelly et al., 2003
SC08	C08	Anthravnose	910 cis	AGA ATG CCT TTA GCT GTT GG CAG AGA GGC TAG GCT TAT CG	<i>Co-4</i>	B8*	Queiroz et al., 2004b *Kelly et al., 2003
SAS13	AS13	Anthravnose	950 cis	CAC GGA CCG AAT AAG CCA CCA ACA CAC GGA CCG AGG ATA CAG TGA AAG	<i>Co-4</i> ²	B8*	Young et al., 1998 *Kelly et al., 2003
SH18	H18	Anthravnose	1100 cis	CCA GAA GGA GCT GAT AGT ACT CCA CAA C GGT AGG CAC ACT GAT GAA TCT CAT GTT GGG	<i>Co-4</i> ²	B8*	Awale and Kelly, 2001 *Kelly et al., 2003
SBB14	BB14	Anthravnose	1150/1050 codominant	GTG GGA CCT GTT CAA GAA TAA TAC GTG GGA CCT GGG TAG TGT AGA AAT	<i>Co-4</i> ²	B8*	Awale and Kelly, 2001 *Kelly et al., 2003
SAB3	AB-3	Anthravnose	400 cis	TGG CGC ACA CAT AAG TTC TCA CGG TGG CGC ACA CCA TCA AAA AAG GTT	<i>Co-5</i>	B7	Vallejo and Kelly, 2001 Campa et al., 2005

SZ20	Z20	Anthrachnose	845 cis	ACC CCT CAT GCA GGT TTT TA CAT AAT CCA TTC ATG CTC ACC	<i>Co-6</i>	B7*	Queiroz et al., 2004b *Kelly et al., 2003
SZ04	Z04	Anthrachnose	567 trans	GGC TGT GCT GAT TAA TTC TGG TGC TCA TTT TAT AAT GGA GAA AAA	<i>Co-6</i>	B7*	Queiroz et al., 2004b *Kelly et al., 2003
SB12	B-12	Anthrachnose	350 cis	CCT TGA CGC ACC TCC ATG TTG ACG ATGGG TTG GCC	<i>Co-9</i>	B4	Mendez de Vigo et al., 2002
SF10	F10	Anthrachnose	1072 cis	GGA AGC TTG GTG AGC AAG GA GGA AGC TTG GCT ATGATG GT	<i>Co-10</i>	B4	Correa et al., 2000 Alzate-Marin et al., 2003
SH13	H13	Angular leaf spot ALS	520 cis	GAC GCC ACA CCC ATT ATG TT GCC ACA CAG ATG GAG CTT TA	<i>Phg-1</i>		Queiroz et al., 2004a
SN02	N02	ALS	890 cis	ACC AGG GGC ATT ATG AAC AG ACC AGG GGC AAC ATA CTA TG	<i>Phg-2</i>	B8*	Nietsche et al., 2000 *Miklas, PC, 2002

	E-ACA/M-CTT ₃₃₀	ALS	280 / 305 codominant	CTT GTT CTG AGT CAT TTA CCT TGC GAA TTC ACA GTC CAA ACT CTA ATC	G 10474 Dominant gene		Mahuku et al., 2004
SAA19	AA19	ALS	650 cis	TGA GGC GTG TCA ATG GAT ATA A GAG GCG TGT TGA TAA TTC TGG	Ouro Negro dominant gene		Queiroz et al., 2004a
SBA16	BA16	ALS	560 cis	TTC CAC GTC TAT TTT GCA TCA CAC GCA TCA CGC AGA ACT	Ouro Negro dominant gene		Queiroz et al., 2004a
SM02	M02	ALS	460 cis	CAA CGC CTC ATT AAA TTG GA CGC CTC TAA ACG GGA GAA AC	Ouro Negro dominant gene		Queiroz et al., 2004a
SU20	U20	Fusarium wilt	750	ACAGCCCCCATTGTGAATTGTAT ACAGCCCCCACACTTATGGCA	A55	B10	Brick et al., 2006 Fall et al., 2001
SW6-800R	OP W-06	Bean pod weevil, <i>Apion godmani</i>	520 cis	AGGCCCATGCCCCCTTAT TGGAGTCGGTCAAACCCATGTT	Agm? J-117	B11	Blair et al., 2006

SCAR Name	PCR	Protocol
SAP6	34 cycles of 10s at 94°C, 40s at 55°C, and 120s at 72°C; followed by one cycle of 5 minutes at 72°C	
BC409	34 cycles of 10s at 94°C, 60s at 70°C, and 120s at 72°C; followed by one cycle of 5 minutes at 72°C	
SU91	34 cycles of 10s at 94°C, 40s at 58°C, and 120s at 72°C; followed by one cycle of 5 minutes at 72°C	
BC420/LG5	35 cycles of 30s at 94°C, 30s at 50°C, and 60s at 72°C; followed by one cycle of 5 minutes at 72°C	
R7313	34 cycles of 10s at 94°C, 40s at 60°C, and 120s at 72°C; followed by one cycle of 5 minutes at 72°C	
R4865	34 cycles of 10s at 94°C, 40s at 60°C, and 120s at 72°C; followed by one cycle of 5 minutes at 72°C	
SR13	95°C 5 min for 1 cycle, 30 cycles of 60s at 94°C, 60s at 60°C, and 60s at 72°C; followed by one cycle of 7 minutes at 72°C	
STB	95°C 5 min for 1 cycle, 30 cycles of 10s at 94°C, 40s at 65°C, and 120s at 72°C; followed by one cycle of 7 minutes at 72°C	
SH11	95°C 5 min for 1 cycle, 30 cycles of 10s at 94°C, 40s at 67°C, and 120s at 72°C; followed by one cycle of 7 minutes at 72°C	
SB10	94°C 5 min for 1 cycle, 30 cycles of 10s at 94°C, 40s at 65°C, and 120s at 72°C; followed by one cycle of 5 minutes at 72°C	
SW13	34 cycles of 10s at 94°C, 40s at 67°C, and 120s at 72°C; followed by one cycle of 5 minutes at 72°C	
ROC11	34 cycles of 10s at 94°C, 40s at 55°C, and 120s at 72°C; followed by one cycle of 5 minutes at 72°C	
SBD5	34 cycles of 10s at 94°C, 40s at 65°C, and 120s at 72°C; followed by one cycle of 5 minutes at 72°C	
SR2	34 cycles of 10s at 94°C, 40s at 60°C, and 120s at 72°C; followed by one cycle of 5 minutes at 72°C 60°C annealing = codominant; 65°C annealing = dominant	
SW12	1 cycle of 60s at 94°C; 30 cycles of 30s at 94°C, 30s at 70°C, and 60s at 72°C; followed by one cycle of 5 minutes at 72°C	
SAS8	1 cycle of 5 min at 94°C; 30 cycles of 60s at 94°C, 60s at 68°C, and 60s at 72°C; followed by one cycle of 7 mins at 72°C	
SK14	34 cycles of 10s at 94°C, 40s at 63°C, and 120s at 72°C; followed by one cycle of 5 minutes at 72°C	
SA14	1 cycle of 5 min at 94°C; 35 cycles of 60s at 94°C, 60s at 55°C, and 90s at 72°C; followed by one cycle of 5 minutes at 72°C	
SI19	34 cycles of 10s at 94°C, 40s at 67°C, and 120s at 72°C; followed by one cycle of 5 minutes at 72°C	
SBC6	94°C 2 min; 30 cycles of 30s at 94°C, 60s at 59 or 65°C?, and 120s at 72°C; followed by one cycle of 7 minutes at 72°C	
SAD12	Annealing temperature of 71°C	
SAE19	94°C 5 min; 35 cycles of 15s at 94°C, 60s at 58°C, and 90s at 72°C; followed by one cycle of 7 minutes at 72°C	
UR11-GT2	60°C annealing = codominant; 65°C annealing = dominant	

KB126	1 cycle 94C for 5 min; 35 cycles at 94C 1 min, 45C 1 min and 72_C for 1 min, final elongation step of 5 min at 72 ⁰ C
SF10	1 cycle of 94 ⁰ C of 3 min: 35 Cycles at 15s at 94 ⁰ C, 60s at 65 ⁰ C and 90s at 72 ⁰ C; followed by 1 cycle of 7 min at 72 ⁰ C
SBA8	1 cycle of 94 ⁰ C of 3 min; 35 Cycles at 15s at 94 ⁰ C, 60s at 65 ⁰ C and 90s at 72 ⁰ C; followed by 1 cycle of 7 min at 72 ⁰ C
Phs	34 cycles of 10s at 94 ⁰ C, 40s at 50 ⁰ C, and 120s at 72 ⁰ C; followed by one cycle of 5 minutes at 72 ⁰ C
SAU5	34 cycles of 10s at 94 ⁰ C, 40s at 60 ⁰ C, and 120s at 72 ⁰ C; followed by one cycle of 5 minutes at 72 ⁰ C
SS18	34 cycles of 10s at 94 ⁰ C, 40s at 63 ⁰ C, and 120s at 72 ⁰ C; followed by one cycle of 5 minutes at 72 ⁰ C
SQ4	34 cycles of 10s at 94 ⁰ C; 40s at 59 ⁰ C, 2 min at 72 ⁰ C, followed by one cycle of 5 min at 72 ⁰ C
SCAreoli	58 ⁰ C annealing temperature followed by <i>DraI</i> digestion
SY20	35 cycles of 30s at 94 ⁰ C, 60s at 65 ⁰ C, and 90s at 72 ⁰ C
SC08	35 cycles of 30s at 94 ⁰ C, 60s at 65 ⁰ C, and 90s at 72 ⁰ C
SAS13	34 cycles of 10s at 94 ⁰ C, 144s at 72 ⁰ C; followed by one cycle of 5 minutes at 72 ⁰ C
SH18	34 cycles of 10s at 94 ⁰ C, 40s at 62 ⁰ C, and 120s at 72 ⁰ C; followed by one cycle of 5 minutes at 72 ⁰ C
SBB14	34 cycles of 10s at 94 ⁰ C, 40s at 67 ⁰ C, and 120s at 72 ⁰ C: followed by one cycle of 5 minutes at 72 ⁰ C
SAB3	1 cycle of 3 mins at 94 ⁰ C; 30 cycles of 10s at 94 ⁰ C, 30s at 65 ⁰ C, 2 minutes at 72 ⁰ C; followed by 1 cycle of 5 mins at 72 ⁰ C
SZ20	35 cycles of 30s at 94 ⁰ C, 60s at 60 ⁰ C, and 90s at 72 ⁰ C
SZ04	45 cycles of 30s at 94 ⁰ C, 120s at 45 ⁰ C, and 90s at 72 ⁰ C
SB12	1 cycle of 94 ⁰ C of 2 min: 35 Cycles of 60s at 94 ⁰ C, 60s at 68 ⁰ C, and 60s at 72 ⁰ C; followed by 1 cycle of 7 min at 72 ⁰ C
SF10	1 cycle of 94 ⁰ C of 3 min: 35 Cycles at 15s at 94 ⁰ C, 60s at 65 ⁰ C and 90s at 72 ⁰ C; followed by 1 cycle of 7 min at 72 ⁰ C
SU20	30 Cycles at 60s at 94 ⁰ C, 30s at 70 ⁰ C and 60s at 72 ⁰ C
SH13	35 cycles of 30s at 94 ⁰ C, 60s at 59 ⁰ C, and 90s at 72 ⁰ C
SN02	30 Cycles of 30s at 94 ⁰ C, 60s at 65 ⁰ , and 90s at 72 ⁰ C
G10474	1 cycle of 94 ⁰ C of 5 min: 35 Cycles of 30s at 94 ⁰ C, 45s at 60 ⁰ C, and 30s at 72 ⁰ C; followed by 1 cycle of 10 min at 72 ⁰ C
SAA19	35 cycles of 30s at 94 ⁰ C, 60s at 56 ⁰ C, and 90s at 72 ⁰ C
SBA16	35 cycles of 30s at 94 ⁰ C, 60s at 58 ⁰ C, and 90s at 72 ⁰ C
SM02	35 cycles of 30s at 94 ⁰ C, 60s at 58 ⁰ C, and 90s at 72 ⁰ C

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